Morphological variation of cinnamon (Cinnamomum verum Persl) germplasm in Matara District of Sri Lanka

Rumana Azad¹, R.A.A.K. Ranawaka², Gamini Senanayake³, K.L. Wasantha Kumara³, D.K.N.G. Pushpakumara⁴, K.G.G. Wijesinghe⁵ and Sudarshanee Geekiyanage³*

¹Board of Study in Agriculture, Faculty of Graduate Studies, University of Ruhuna, Matara, Sri Lanka, ²Mid Country Research Station, Department of Export Agriculture, Sri lanka,

³Department of Agricultural Biology, Faculty of Agriculture, University of Ruhuna, Mapalana, Kamburupitiya, Sri Lanka,

⁴Department of Crop Science, Faculty of Agriculture, University of Peradeniya, Peradeniya, Sri Lanka ⁵National Cinnamon Research and Training Center, Department of Export Agriculture, Palolpitiya, Thihagoda, Sri Lanka

*Email: sudarshanee@agbio.ruh.ac.lk

ABSTRACT

Forty seven representative cinnamon accessions were collected from Matara District of Sri Lanka to analyze the morphological variation of *Cinnamomum verum* germplasm. Morphological characters viz. Leaf length, Leaf width, Leaf length-width ratio, Petiole length, Leaf arrangement, Leaf shape, Leaf apex, Leaf base, Leaf texture, Upper surface leaf color, Flush color, Bark color, Bark surface, and Bark fragrant were recorded. Principal component analysis (PCA) using four quantitative morphological characteristics, indicated that the first two principal components (PCs) with Eigen values of more than one and accounted for 88.88% of the total variance. Cluster analysis classified 47 accessions into nine groups. The present study demonstrates a considerable diversity of morphological characters among the accessions that can be useful in germplasm management and future crop improvement programs.

Key words: Cinnamomum verum germplasm, morphological characters, Principal component analysis, Cluster analysis

INTRODUCTION

The genus Cinnamomum belongs to the family Lauraceae and consists of about 110 species of evergreen trees and shrubs (Purseglove et al., 1969). Out of nine Cinnamomum species, Cinnamomum verum is one of the most important species in Sri Lanka which contributes to 70% of the world true cinnamon bark production (Abeysinghe et al., 2009). The export volume of cinnamon and earning for 2012 was 14,435 metric tons and 16,654.7 million rupees respectively (Sri Lanka Custom, 2012). Cinnamon is endemic to Sri Lanka and it had been found in central hilly areas of Sri Lanka and also in Sinharaja and Knuckles forest reserves. The ideal environmental condition for cinnamon is available in wet zone of Sri Lanka. However commercial cinnamon cultivation is carried out in intermediate zones of mid and low country, where annual rainfall is more than 1750 mm. Cinnamon is mostly cultivated along the coastal belts of Kalutara, Galle, Hambantota at an elevation of about 250 m above mean sea level. The most suitable temperature is between 25°C- 32°C (Department of Export Agriculture, 2013). Cinnamon bark is mainly used as a spice for flavoring food product and leaf oil is used as flavor ingredients and also in cosmetics and pharmaceutical industries (Paranagama et al., 2001). Different biological activities including anti-diabetic, anti-inflammatory, astringent and diuretic effects have

been popularized cinnamon in folk medicine (Lee *et al.*, 2010). In modern medicine, cinnamon is combined with other ingredients to treat diarrhea, internal hemorrhage, impotency, typhoid, halitosis, checking nausea and vomiting and for restoring normal skin color on the face (Warrier *et al.*, 1994).

According to Ravindran *et al.*, (2004) leaf shape of cinnamon varies from oval or elliptic to lanceolate-oval or narrowly elliptic, $3 \times 7 - 8 \times 25$ *cm, leaf apex shortly or broadly acuminate and leaf base acutish or cuneate.* A study by Wijesinghe and Gunarathna, (2001) showed correlation between leaf size and shape with yield in seven different types of true cinnamon. According to this observation trees with large round leaves and big leaves had high bark yield. Moreover bark oil (cinnamaldehyde %) quality is higher in the variety of inwardly curved leaves and high quality leaf oil was obtained from the small round leaves.

Cinnamon flower exhibits protogynous dichogamy and it is cross pollinated (Joseph, 1981). Thus, vegetative propagation is necessary for producing uniformly high yielding populations and for propagating elite lines (Rema *et al.*, 1997). A core collection is a representative subset of a large number of populations which intends to improve management

and use of a germplasm collection (Diwan *et al.*, 1995). It is also a powerful material for evaluation of germplasm, identification of trait-specific accessions, gene discovery, allele mining, genomic study, marker development, and molecular breeding (Qiu *et al.*, 2013). Cluster analysis followed by Principal component analysis had been used to cluster *Cinnamomum* spp. into groups and to show relationship among the species on the basis of morphological characters (Ravindran *et al.*, 1991).

Morphological variation of a crop indicates the genetic diversity and effect from environment. Both environmental and genetic effects contribute to phenotypic variation within and among populations (Allard and Bradshaw, 1964; Andrew et al., 2010). Some molecular study has been conducted to evaluate genetic differences on Cinnamomum species (Ho and Hung, 2011; Joy and Maridass, 2008; Lin et al., 1997; Kameyama, 2012; Soulange et al., 2007; Lee et al., 2010; Kojoma et al., 2002; Sandigawad and Patil, 2011; Kuo et al., 2010). A molecular research has been done on the genetic analysis of Cinnamomum species by sequencing TrnL intron region, intergenic spacer between trnT-trnL, trnL-trnF, trnH-psbA and nuclear ITS (Abeysinghe et al., 2009). Another work has been carried out to find a more reliable approach to identify Cinnamomum species correctly using RAPD and SRAP techniques. Some primers gave highly polymorphic banding patterns using these techniques. This preliminary study showed that using these molecular markers, it is possible to identify the Cinnamomum species (genus specific and species specific) and intra-species variations (Abeysinghe et al., 2014). Therefore, the present study is focused on the Cinnamomum germplasm which were collected from different locations of Matara district to analyze the morphological variation.

MATERIALS AND METHODS

Total forty seven accessions were collected according to their distinct morphological characters from these cinnamon growing areas particularly from Deiyandara, Ehala Athuraliya, Karapotu Gala, Palolpitiya, Ehalawitiyala and Kamburupitiya during September, 2014. Semi-hard 1/1.5 inch stem cutting with 1 or 2 leaves and active buds from every accession had been planted in nursery for further studies. In the Laboratory the Leaf length (LL), Leaf width (LW), Leaf length-width ratio (LLWr), Petiole length (PL) of samples were measured while other morphological traits Leaf arrangement (LA), Leaf shape (LS), Leaf apex (LAP), Leaf base (LB), Leaf texture (LT), Upper surface leaf color (ULC), Flush color (FC), Bark color (BC), Bark surface (BS), and Bark fragrant (BF) were observed in the time of every field visit during September, 2014. Flush color had been observed following Munsell Color Chart (Munsell Color, 1977). Morphological analysis of collected samples was done considering both quantitative and qualitative characters.

Analysis of variance applying descriptive statistics such as mean, standard deviation, coefficient of variation and correlation coefficient for quantitative traits were calculated. Principal Component Analysis (PCA) was conducted in order to identify the patterns of morphological variation using IBM SPSS Statistics 20.0 software (version 20), IBM, USA. Clustering of genotypes into similar groups was carried out using Ward's hierarchical algorithm based on squared Euclidean distances.

RESULTS AND DISCUSSION

Fourteen morphological characters were recorded from 47 accessions. Among fourteen morphological characters, four quantitative characters of Leaf length (LL), Leaf width (LW), Leaf lengthwidth ratio (LLWr), Petiole length (PL) and ten qualitative characters of Leaf arrangement (LA), Leaf shape (LS), Leaf apex (LAP), Leaf base(LB), Leaf texture (LT), Upper surface leaf color (ULC), Flush color (FC), Bark color (BC), Bark surface (BS), and Bark fragrant (BF) were recorded.

The correlation coefficient was observed between four different morphological traits (Table 1). There were positive significant linear relationships between the Leaf width and Leaf length (0.699), Petiole length and Leaf length (0.613) and Leaf width and Petiole length (0.574) at 0.01% significant level. The positive and significant relationships among the traits will provide plant breeders an understanding on manipulation of such traits.

Variable	LL	LW	LLWr	PL
LL	-	.699**	.367*	.613**
LW		-	396**	.574**
LLWr			-	.045
PL				-

Table 1. The correlation of quantitative traits according to Pearson's Correlation Coefficient evaluated in cinnamon germplasm collections.

 \ast Correlation is significant at the 0.05 level (2-tailed).

** Correlation is significant at the 0.01 level (2-tailed).

The Principal Component Analysis showing the factor scores of each character among the cinnamon accessions, eigan values and percentage total variance accounted by four principal component (Table 2). The PCA was used to remove redundancy in the data set. The first two principal components (PC-1 and PC-2) accounted for most of the variability observed among the accessions and their eigen value is more than 1. The first two principal components accounted for 88.88% of the total variability where PC-1 explained 56.45% of the total variability was loaded on LL, LW and PL and the PC-2 accounted for 32.43% of the variation and was loaded on LLWr. PCA is commonly used to analyze large data sets. It has been used to evaluate germplasm of rice (Sohrabi *et al.*, 2012), olive (Cantini *et al.*, 1999), vineyard peach (Nikolic *et al.*, 2010), peach (Perez *et al.*, 1993), loquat (Leguizamón *et al.*, 2003; Badenes *et al.*, 2000; Martinez-Calvo *et al.*, 2008), and apricot (Yilmaz *et al.*, 2012; Ruiz and Egea, 2008). Its main purpose is to extract the important information from the table, to represent it as a set of new orthogonal variables called principal components, and to show the pattern of similarity of the observations and of the variables as points in maps (Abdi and Williams, 2010).

Table 2. Eigen values of the correlation matrix and their contribution to total variation of cinnamon germplasm collections.

Component	Ir	iitial Eigenva	lues	Extrac	tion Sums of Loadings	f Squared	Rota	ation Sums of Squared Loadings		
	Total	% of Variance	Cumulati ve %	Total	% of Variance	Cumulati ve %	Total	% of Variance	Cumulativ e %	
1	2.25	56.47	56.47	2.25	56.47	56.47	2.25	56.45	56.45	
2	1.29	32.40	88.88	1.29	32.40	88.88	1.29	32.43	88.88	
3	.43	10.92	99.80							
4	.008	.193	100.00							

The cluster analysis using Ward's method classified the 47 cinnamon accessions in to nine clusters at rescaled distance of 3.0 (Fig. 1). Accessions from Cluster no. 8 showed highest CV% of LL (15.10%), LW (13.53%) and PL (28.28%), which indicated those accessions are highly variable in contrast to the other accessions (Table 3). On the other hand, CV% of LW (11.61%) and PL (17.30%)

of the accessions from cluster no. 9 also higher than the other accessions. Accessions belong to cluster no. 1 and cluster no. 5 showed variation in CV% of PL (11.73% and 22.12% respectively) comparing other accessions. All accessions in one cluster represent the respective cluster in terms of qualitative characters while the qualitative characters vary among clusters.

Table 3. Mean (M), Standard Deviation (SD) and Coefficient of Variation (CV%) of quantitative characters according to clusters.

		LL LW LLWr PL				PL						
Cluster number	М	SD	CV %	М	SD	CV%	М	SD	CV %	М	SD	CV%
1	14.01	1.22	8.73	6.51	0.36	5.55	2.15	0.12	5.71	2.06	0.24	11.73
2	18.70	0	0	9.13	0	0	2.05	0	0	2.10	0	0
3	12.90	0.63	4.91	7.68	0.03	0.45	1.68	0.09	5.15	1.90	0.17	9.12
4	19.48	0.59	3.03	7.44	0.68	9.14	2.63	0.17	6.43	2.37	0.25	10.63
5	11.37	0.91	7.97	5.73	0.50	8.67	1.99	0.09	4.61	1.33	0.29	22.12
6	7.43	0	0	3.40	0	0	2.19	0	0	1.20	0	0
7	12.64	1.04	8.22	4.73	0.33	7.09	2.67	0.08	2.87	1.36	0.13	9.38
8	15.45	2.33	15.10	5.75	0.78	13.53	2.68	0.04	1.60	2.00	0.57	28.28
9	13.37	1.28	9.60	5.71	0.66	11.61	2.35	0.09	3.66	1.67	0.29	17.30



Figure 1: Dendogram of cinnamon accessions derived through Ward's linkage Cluster Analysis based on four quantitative traits.

The qualitative characters of all the accessions were arranged in Table-5 following quantitative characters originated from nine clusters. Qualitative characters of LA, LS, LB and ULC of the accessions belong to cluster 4 and cluster 8 are similar to each other whereas, cluster 1, 5, 7 and 9 with large number of accessions showed considerable variation in qualitative traits. Flush color of cinnamon accessions showed a variation in the collection. There may be a relationship between their oil contents and flush color as previously reported by (Krishnamoorthy et al., 1988). Accessions from all the nine clusters showed

variations in Leaf shape (Fig. 2). One accession (MPT-03) belongs to cluster no.03 have ovate Leaf shape which is different from the other accessions. In addition, accessions MEA-11 and MKG-01 grouped in to cluster no.07 and cluster no.08 respectively showed narrowly elliptic Leaf shape which is not similar to the other accessions. According to the two types of trait combination it has been clearly manifested that both types of traits has a substantial contribution in accession grouping. These variations ensured genetic differentiation and allele richness among the accessions.

Table 4. Qualitative traits with Type code	in cinnamon germplasm.
--------------------------------------------	------------------------

S. No.	Qualitative traits	Pattern/ Color type	Type code
1	Leaf arrangement (LA)	Opposite	1
		Sub-opposite Opposite or	2
		sub-opposite Opposite to	3
		sub-opposite	4
2	Leaf shape (LS)	Filiptic	1
2	Leaf shape (LS)	Langeolete	2
		Quete	23
		Norrowly alliptic Elliptic	1
		to broadly alliptic Quete	5
			7
		Overte oblong to overte langeolete	11
		Ovale-oblong to ovale-nanceonale	11
3	Leaf apex (LAP)	Acute	1
	-	Obtuse	2
		Acuminate	3
		Gradually acuminate	4
		Long acuminate	5
		Broadly acuminate	6
		Narrowly acuminate	7
		Blunt or subacute	10
Α	Leefher (LD)	A system Of the set	1
4	Leaf base(LB)	Acute Obluse	1
		Cuneate	2 3
		Shortiy acute	5
		Rounded of subacute	5
		Coriacerous	0
5	Leaf texture (LT)	Subcoriaceous	1
		Rigidly coriaceous	2
		Thinly to stiffly coriaceous	3
		Chartaceous to rigidly chartaceous	4
		Chartaceous	5
			6
		Dark green	
6	Upper surface leaf color (ULC)	Green	1
			3
		2.5R 7/6	
7	Flush color (FC)	2.5R 6/8	2
		5GY 7/10	4
		2.5GY 8/10	6
		2.5GY 8/6	7
		2.5R 4/8	8
		2.5R 7/8	9
			12
		Brown Whitish	1
8	Bark color (BC)	brown Light	
		brown	2
			3
_		Slightly rough	1
9	Bark surface (BS)	Kough	1 2
		Smooth	2 1
		Weak fragment ereme	7
10		weak iragrani aroma	1
10	Bark Iragrant (BF)	Cood from the second	2
		Strong fragment aroma	3
		Strong fragrant aroma	4
1			'

Cluster	Accession	Qualitative characters									
number	number	LA	LS	LAP	LB	LT	ULC	FC	BC	BS	BF
	MEA 12	3	5	3	1	5	1	9	1	2	1
	MKG 8	4	11	4	5	5	1	7	3	2	2
	MEA 6	2	1	1	5	1	1	9	1	1	2
	MEA 2	3	5	6	5	6	3	9	3	4	1
	MPE 8	4	2	6	1	5	1	12	1	4	3
1	MEA 3	4	7	7	5	5	1	12	1	1	2
	MEA 4	4	5	10	1	6	1	4	1	1	2
	MKG 11	1	1	1	1	3	1	12	2	1	1
	MPT 1	4	7	3	6	6	1	2	1	1	2
	MMK 18	4	2	1	1	6	1	12	3	4	1
	MKG 15	2	2	6	1	6	3	8	2	1	2
2	MMK 20	2	5	1	5	5	1	6	3	4	2
	MEA 1	4	7	1	5	6	1	2	1	1	2
3	MKG 9	4	5	10	5	6	1	9	1	4	2
	MPT 3	4	3	2	2	1	1	9	1	4	2
	MPT 4	4	2	3	1	5	1	12	1	4	4
4	MMK 13	4	2	5	1	6	1	6	1	4	4
	MKG 12	4	2	3	1	6	1	9	3	4	1
	MKG 2	1	1	10	6	3	3	9	3	4	1
	MKG 7	1	7	10	6	2	1	6	3	4	1
	MPT 5	4	7	3	2	1	1	9	3	4	2
	MMK 11	2	5	1	1	1	3	6	3	4	3
	MMK 15	4	5	4	1	5	1	12	1	4	2
5	MPE 10	2	1	3	6	6	1	12	3	4	2
	MMK 17	4	1	1	1	4	1	6	1	1	2
	MKG 6	1	1	4	6	1	3	7	1	1	1
	MKG 13	4	1	3	5	5	1	4	2	1	1
	MKG 5	1	7	10	6	3	1	4	1	4	2
	MEA 5	4	7	3	6	5	1	7	2	2	2
6	MMK 14	2	1	10	5	6	1	7	1	4	3
	MEA 11	2	4	1	1	4	1	12	1	2	1
	MKG 10	4	4	10	5	6	1	9	1	4	2
	MEA 10	3	4	10	1	6	3	8	3	1	3
7	MPE 7	4	1	7	1	5	1	7	1	4	4
	MPT 2	4	2	3	1	5	1	9	1	4	3
	MPE 9	2	4	3	3	6	1	7	1	4	2
	MKG 4	4	2	3	1	6	1	9	2	2	1
8	MKG 1	4	4	3	1	6	1	2	2	4	2
	MMK 16	4	4	5	1	6	1	4	1	1	1
	MPE 6	4	1	3	1	6	1	12	1	1	2
	MMK 19	4	1	7	1	2	1	4	1	4	2
	MEA 8	4	2	4	1	3	1	9	1	2	1
9	MKG 3	1	7	3	1	2	3	9	3	4	1
	MMK 12	2	1	4	1	6	1	9	3	4	2
	MEA 7	3	1	3	5	6	1	9	3	2	1
	MEA 9	3	5	4	5	5	1	7	3	2	2
	MKG 14	2	1	4	1	6	1	12	2	1	4

 Table 5. Variation of qualitative characters within clusters which were derived through Ward's linkage method.



Figure 2: Variation in leaf shape within clusters; Cluster-01: 01(MEA-06) Elliptic, 02 (MMK-18) Lanceolate, 03 (MEA-02) Elliptic to broadly elliptic, 04 (MPT-01) Ovate-lanceolate, 05 (MKG-08) Ovateoblong to ovate-lanceolate; Cluster-02: 06 (MMK-20) Elliptic to broadly elliptic; Cluster-03: 07 (MPT-03) Ovate, 08 (MKG-09) Elliptic to broadly elliptic, 09 (MEA-01) Ovate-lanceolate; Cluster-04: 10 (MMK-13) Lanceolate ; Cluster-05: 11 (MKG-06) Elliptic, 12 (MMK-11) Elliptic to broadly elliptic, 13 (MEA-05) Ovate-lanceolate; Cluster-06: 14 (MMK-14) Elliptic; Cluster-07: 15 (MPE-07) Elliptic, 16 (MPT-02) Lanceolate, 17 (MEA-11) Narrowly elliptic; Cluster-08: 18 (MKG-01) Narrowly elliptic; Cluster-09: 19 (MMK-12) Elliptic, 20 (MEA-08) Lanceolate, 21 (MEA-09) Elliptic to broadly elliptic, 22 (MKG-03) Ovate-lanceolate.

There is a variation in qualitative and quantitative characters among the cinnamon accessions of Matara district which were categorized into nine distinct clusters at rescale distance of 3. Accessions MMK14 and MMK 20 could be distinct

REFERENCES

- Abdi, H. and Williams, L.J. 2010. Principal component analysis. *Wiley Interdisciplinery Review: Comput. Stat.*, **2**:433-459.
- Abeysinghe, P.D., Samarajeewa, N.G.C.D., Li, G., and Wijesinghe, K.G.G. 2014. Preliminary investigation for the identification of Sri Lankan *Cinnamomum* species using randomly amplified polymorphic DNA (RAPD) and sequence related amplified polymorphic (SRAP) markers. J. National Sci. Foundation Sri Lanka, **42**(3): 175-182.
- Abeysinghe, P.D., Wijesinghe, K.G.G., Tachida, H. and Yoshda, Τ. 2009. Molecular characterization of cinnamon (CinnamomumverumPresl.) accessions and evaluation of genetic relatedness of cinnamon species in Sri Lanka based on TrnL intron region, intergenic spacer between trnT-trnL, trnL-trnF, trnH-psbA and nuclear ITS. Research Journal of Agriculture and Biological Sciences, 5(6): 1079-1088.
- Allard, R.W. and Bradshaw, A. 1964. Implications of genotype-environmental interactions in applied plant breeding. *Crop Science*, **4**(5): 503-508.
- Andrew, R.L., Wallis, I.R., Harwood, C.E. and Foley, W.J. 2010. Genetic and environmental contributions to variation and population divergence in a broad-spectrum foliar defence of *Eucalyptus tricarpa.Ann. Bot.* **105**:707-717.
- Badenes, M.L., Martínez-Calvo, J. and Llacer, G. 2000. Analysis of a germplasm collection of loquat (*Eryobotria japonica*Lindl.). *Euphytica*, **114**:187-194.
- Cantini, C., Cimato, A. and Sani, G. 1999. Morphological evaluation of olive germplasm

genotypes they clustered independently. as Significant positive correlations were observed between Leaf width and Leaf length (0.699), Petiole length and Leaf length (0.613) and Leaf width and Petiole length (0.574). Morphological diversity among accessions would indicate the genetic diversity which could be used for exploitation of economically important traits of cinnamon in the future. The next step of this research project through molecular and chemical characterization of representative accessions from each cluster would provide precise information on economically important genotypes and the morphological markers for better genotypes.

ACKNOWLEDGEMENTS

Authors wish to acknowledge the project Transforming University of Ruhuna into International Status (TURIS) 2013 grant for financial support and Mrs. Sheron Weerasooriya from Cinnamon Research Station and Extension Officers of Department of Export Agriculture (DEA) for assistance in germplasm collection.

present in Tuscany region. *Euphytica*, **109**:173-181.

- Department of Export Agriculture, Sri Lanka. 2013. [Online] [Accessed on 27th July 2015]http://www.exportagridept.gov.lk/web/in dex.php?option=com_content&view=article&i d=128&Itemid=159&Iang=en
- Diwan, N., McIntosh, M.S. and Bauchan, G.R. 1995. Methods of developing a core collection of annual Medicago species. Theoretical and Applied Genetics, 90(6): 755-761.
- Joy, P. and Maridass, M. 2008. Inter species relationship of Cinnamomum species using RAPD marker analysis. Ethnobotanical Leaflets, 12: 476 - 480.
- Joseph, J.1981. Floral biology and variation in cinnamon. PLACROSYM IV, pp. 431 434.
- Ho, K.Y. and Hung, T.Y. 2011. Cladistic relationships within the genus Cinnamomum (Lauraceae) in Taiwan based on analysis of leaf morphology and inter-simple sequence repeat (ISSR) and internal transcribed spacer (ITS) molecular markers. Afr. J. Biotechnol., 10: 4802 - 4815.
- Kameyama, Y. 2012. Development of microsatellite markers for Cinnamomumcamphora (Lauraceae). Am. J. Bot., 99(1): e1 – e3.
- Kuo, D.C., Lin, C.C., Ho, K.C., Cheng, Y.P., Hwang, S.Y. and Lin, T.P. 2010. Two genetic divergence centers revealed by chloroplastic DNA variation in populations of CinnamomumkanehiraeHay. Conservation Genetics, 11: 803-812.
- Kojoma, M., Kurihara, K., Yamada, K., Sekita, S., Satake, M. and Lida, O. 2002. Genetic identification of cinnamon (Cinnamomum spp.) based on the trnL-trnF chloroplast DNA. Planta Med., 68: 94-96.

- Krishnamoorthy, B., Gopalam, A. and Abraham, J. 1988. Quality parameters of cinnamon (Cinnamomumverum) in relation to flush colour. Indian Cocoa Arecanut Spices J., 12: 38.
- Lee, S.C., Chiou, S.J., Yen, J.H., Lin, T.Y., Hsieh, K.T.and Yang, J.C. 2010. DNA barcoding Cinnamomum osmophloeum Kaneh. based on the partial non-coding ITS2 region of ribosomal genes. J. Food Drug Anal., 18(2): 128-135.
- Lee, S.C., Lee, C.H., Yilin, M. and Ho, K.Y. 2010. Genetic identification of Cinnamomum species based on partial internal transcribed spacer 2 of ribosomal DNA. J. Food and Drug Anal., 18(4): 225-231.
- Leguizamón, J. and Badenes, M.L. 2003. Multivariate analysis as a tool for germplasm studies, example of analysis of germplasm loquat data. Acta Hort., 606:29-35
- Lin, T.P., Cheng, Y.P. and Huang, S.G. 1997. Allozyme variation in four geographic areas of Cinnamomum kanehlrae. Journal of Heredity, 88: 433 - 438.
- Martinez-Calvo, J., Gisbert, A.D., Alamar, M.C., Hernandorena, R., Romero, C., Llacer, G., and Badenes., M.L. 2008. Study of a germplasm collection of loquat (Eriobotrya japonica Lindl.) by multivariate analysis. Genet. Resources Crop Evol. 55:695–703.
- Munsell Color. 1977. Munsell Color Charts for Plant Tissues. Munsell Color, Gretag Macbeth, 617 Little Britain Road, New Windsor, NY 12553-6148.
- Nikolic, D., Rakonjac, V., Milatovic, D., and Fotiric, M. 2010. Multivariate analysis of vineyard peach (Prunuspersica L. Batsch.) germplasm collection. Euphytica, 171: 227-234.
- Paranagama, P.A., Wimalasena, S., Jayatilake, G.S., Jayawardena, A.L., Senanayake, U.M. and Mubarak, A.M. 2001. A comparison of essential oil constituents of bark, leaf, root and fruit of cinnamon (Cinnamomum zeylanicum Blume.) grown in Sri Lanka. J. National. Sci. Foundation Sri Lanka, 29(3&4):147-153.
- Perez, S., Montes, S. and Mejia, C. 1993. Analysis of peach germplasm in Mexico. J. Amer. Soc. Hort. Sci., 118:519–524
- Purseglove, J. W. 1969. Lauraceae. In Tropical Crops: Dicotyledons. Vol. 2, 2nd ed. Purseglove J. W. (ed), Longmans Green and Co. ltd. London, U.K. pp. 187-192.

- Qui, L.J., Xing, L.L., Guo, Y., Wang, J. and Chang, R.Z. 2013. A platform for soybean molecular breeding: the utilization of core collections for food security. Plant Mol. Biol., 83: 41-50.
- Ravindran, P.N., Nirmal-Babu, K. and Shylaja, M. eds. 2004. Cinnamon and Casssia: The Genus Cinnamon. Medicinal and Aromatic plants-Industrial Profiles. CRC Press, Florida.
- Ravindran, S., Krishnaswamy, N. R., Manilal, K.S. and Ravindran, P.N. 1991. A cluster analysis study of Cinnamomum from Kerala, India. Feddes Report., 102: 13-21.
- Rema, J., Krishnamoorthy, B., and Mathew, P.A. 1997. Vegetative propagation of major tree spices – a review.Journal of Spices and Aromatic Crops, 6(2): 87-105.
- Ruiz, D. and Egea, J. 2008. Phenotypic diversity and relationships of fruit quality traits in apricot (PrunusarmeniacaL.) germplasm. Euphytica, 163: 143-158.
- Sandigawad, A.M. and Patil, C.G. 2011. Genetic diversity in Cinnamomum zeylanicum Blume. (Lauraceae) using random amplified polymorphic DNA (RAPD) markers. Afr. J. Biotechnol.,10: 3682-3688.
- Sohrabi, M., Rafii, M.Y., Hanafi, M.M., Siti Nor Akmar, A. and Latif, M.A. 2012. Genetic diversity of upland rice germplasm in Malaysia based on quantitative traits. Sci World J, :416291.
- Soulange, J.G., Sanmukhiya, V.M.R. and Seeburrun, S.D. 2007. Tissue culture and RAPD analysis of Cinnamomum camphora and Cinnamomum verum. Biotechnology, 6:239-244.
- Sri Lanka Customs, 2012. Sri Lanka Customs Annual Performance Report 2012. Sri Lanka Customs.
- Warrier, P.K., Nambiar, V.P.K and Ramankutty, C. 1994. Indian Medicinal Plants A Compendium of 500 Species (Vol. II). Orient Longman Ltd., Madras, India.
- Wijesinghe, K.G.G. and Gunarathna, W.D.L. 2001. Characterization of true Cinnamon (Cinnamomum verum Persl) based on leaf morphology and their relationship with yield and quality. Sri Lanka Association for the Advancement of Science. 57th Annual Session. Proc. Part I. pp 42
- Yilmaz, K.U., Kargi, S.P. and Kafkas, S. 2012. Morphological diversity of the Turkish apricot (Prunus armeniaca L.) germplasm in the Irano-Caucasian ecogeographical group. Turk. J. Agric. For., 36:688–694.